Comprehensive Molecular Phylogeny of the Sub-Family Dipterocarpoideae (Dipterocarpaceae) Based on Chloroplast DNA Sequences

Dayananda Thawalama Gamage¹, Morley P. de Silva², Nobuyuki Inomata¹, Tsuneyuki Yamazaki³ and Alfred E. Szmidt^{1*}

¹Department of Biology, Graduate School of Sciences, Kyushu University, Fukuoka 812-8581, Japan

²Department of Botany, Faculty of Science, University of Ruhuna, Sri Lanka ³Research Institute of Evolutionary Biology, 4-28, Kamiyoga 2, Setagaya, Tokyo

(Received 1 December 2005, accepted 20 December 2005)

Dipterocarpoideae, the largest sub-family of well-known plant family Dipterocarpaceae, dominates in South Asian rain forests. Although several previous studies addressed the phylogeny of the Dipterocarpaceae family, relationships among many of its genera from the Dipterocarpoideae sub-family are still not well understood. In particular, little is known about the relationships of the genera Vateriopsis, Stemonoporus, Vateria and inconsistence remains between phylogenetic results and taxonomic classifications of Shorea and Hopea species. We studied molecular phylogeny of the sub-family Dipterocarpoideae using the trnL-trnFspacer, *trnL* intron and the *matK* gene sequences of chloroplast DNA (cpDNA). This study is the first comprehensive phylogeny reconstruction for the sub-family Dipterocarpoideae based on cpDNA, as it includes most genera (14) and a large number of species (79) with most species endemic to Sri Lanka, as well as one species from Seychelles and one species from the genus Monotes from Madagascar. Phylogenetic trees were constructed using the Neighbor Joining (NJ) and Maximum Likelihood (ML) methods using combined set of sequences including all three cpDNA regions. The topologies of the NJ and ML trees were to a certain extent, consistent with the current taxonomy of Dipterocarpoideae based on morphology and with previous molecular phylogenies based on cpDNA. Furthermore, our results provided new evidence regarding the relationships of the following genera: *Vateriopsis* and *Stemonoporus* and about the validity of the previous morphology based classifications of *Shorea* species. In addition, the topology of our trees was consistent with the classification of Shorea species proposed by Maury (1978), Maury-Lechon (1979) and Symington (1943). Finally, our results provided evidence for the affinity of the genus Monotes to Asian Dipterocarpoideae rather than to Tiliaceae and indicated that it is a good candidate for outgroup species for future studies of the former sub-family.

Key words: chloroplast DNA, Dipterocarpoideae, *Doona*, molecular phylogeny, *Monotes*

INTRODUCTION

Dipterocarpaceae is a well-known plant family with approximately 580–680 species (Ashton, 1977; Ashton, 1982; Maury-Lechon and Curtet, 1998). Many members of this family are large forest emergent trees, typically reaching heights of 40–70 m. Their distribution is pan tropical, from northern South America to Africa, Seychelles, Sri Lanka, Philippines, India, China, Thailand, Indonesia and Malaysia with the greatest diversity and abundance in western Malaysia.

The Dipterocarpaceae family is divided into three subfamilies: (i) Monotoideae, with three genera and about 30 species, distributed across Africa, Madagascar and South America, (ii) Pakaraimoideae with a single species

Edited by Fumio Tajima

^{*} Corresponding author. E-mail: aszmiscb@mbox.nc.kyushu-u.ac.jp

Pakaraimaea roraimae found in the Guaianan highlands of South America and *(iii)* Dipterocarpoideae, the largest of the sub-families, with 13 genera and about 470 species (Ashton, 1982), which distribute mainly in South Asian countries such as India, Sri Lanka, Philippines, China, New Guinea, Indonesia, Thailand and Malaysia with the exception of *Vateriopsis seychellarum*, which is endemic to Seychelles.

The phylogenetic position of the genus *Monotes*, which is often placed in the sub-family Monotoideae (e.g., Maury, 1978) is still unclear. Initially, it was associated with the family Tiliaceae (Heim, 1892). Later however, it was moved to the sub-family Monotoideae of the family Dipterocarpaceae (Gilg, 1925). On the other hand, based on morphology, Maury (1978) and Kostermans (1989) treated Monotoideae as a separate family.

The sub-family Dipterocarpoideae can be further divided into two tribes: Dipterocarpeae and Shoreae (Brandis, 1895). The genera of the first tribe (Anisoptera, Cotylelobium, Dipterocarpus, Stemonoporus, Upuna, Vateria, Vateriopsis and Vatica) have valvate sepals in fruits, solitary vessels, scattered resin canals and the basic chromosome number n = 11. The genera of the second tribe (Dryobalanops, Neobalanocarpus, Hopea, Parashorea and Shorea) have imbricate sepals in fruits, grouped vessels, resin canals in tangential bands and basic chromosome number n = 7 (Ashton, 1982; Brandis, 1895; Jong, and Kaur, 1979). However, there is still much controversy regarding the number of genera of the Dipterocarpoideae sub-family, especially in the Shoreae tribe, which varies depending on the author between nine and 19 (Ashton, 1977; Ashton, 1982; Kostermans, 1978; Kostermans, 1982; Kostermans, 1984; Kostermans, 1992; Maury, 1978; Maury-Lechon, 1979; Meijer, and Wood, 1964; Meijer, and Wood, 1976). Perhaps the most controversial is classification of the genus Shorea. Based on embryo and leaf epidermal characters Maury (1978) divided this genus Shorea into the following separate genera: Shorea, Anthoshorea, Rubroshorea, Richetia, Doona, and Pentacme. On the other hand, Ashton (1977), Ashton (1980) and Ashton (1982) included them in a single genus Shorea, which was further divided into 11 sections: Shorea, Pentacme, Neohopea, Richetioides, Anthoshorea, Rubella, Brachypterae, Pachycarpae, Mutica, Ovalis and Doona. Yet another classification was proposed by Symington (1943) who divided the genus Shorea into three separate genera: Shorea, Pentacme and Parashorea. Further, based on wood anatomy he divided it into the four following wood groups: Balau, Red Meranti, White Meranti and Yellow Meranti.

The phylogenetic relationships of Dipterocarpaceae have been studied using distribution, fossil and morphological data by Ashton (1982) and the first phylogeny based on molecular data was reported by Tsumura et al. (1996). Since then, several other phylogenetic studies on Dipterocarpaceae were reported based on chloroplast (cp) DNA sequences (Dayanandan et al., 1999; Gamage et al., 2003; Kajita et al., 1998; Kamiya et al., 1998; Morton et al., 1999) and the nuclear gene PgiC (Kamiya et al., However, previous studies on molecular phylog-2005).eny of the Dipterocarpaceae included either limited number of species (Kajita et al., 1998; Morton et al., 1999; Tsumura et al., 1996) or informative sites (Gamage et al., 2003; Kamiya et al., 1998) or both (Dayanandan et al., 1999). The most recent work by Kamiya et al. (2005) has mainly focused on the relationships of Shorea, Hopea, Neobalanocarpus and Parashorea genera and did not include species from the Dipterocarpeae tribe and species of the Doona genus (Kostermans, 1984; Kostermans, 1992; Maury, 1978; Maury-Lechon, 1979). As a result, phylogenetic placement of many species and genera, which belong to sub-family Dipterocarpoideae is still unclear. In particular, little is known about the relationships of the following genera: Vateriopsis, Stemonoporus and Vateria and species from the Doona genus created by Kostermans (1984), Kostermans (1992) and Maury (1978).

It is therefore necessary to examine a larger number of species representing all genera and distribution areas. The main objective of the present work was to provide comprehensive assessment of phylogenetic relationships among Dipterocarpoideae species from Southeastern Asia. Another objective of our study was to ascertain the placement of the genus Vateriopsis (endemic to Seychelles) and classification of many endemic Sri Lankan species from the tribes Dipterocarpeae and Shoreae. In addition, our aim was to investigate the familial affinity of the genus Monotes. Finally, we wanted to determine, which of the two species from outside the Dipterocarpoideae sub-family included in our study (Tilia kuisiana or Monotes madagascariensis) is a better candidate for outgroup species for future studies of Dipterocarpoideae phylogeny.

Among the 79 Dipterocarpaceae species included in our present study, 42 species were from Malaysia, 34 species were from Sri Lanka, one species was from Thailand, one species was from Seychelles and one species was from Madagascar. The species used here represent 14 genera of the family Dipterocarpaceae and thus provide the first comprehensive material for phylogeny reconstruction. To address the issues of outgroup choice and the family placement of the genus *Monotes*, we have also included one species from the Tiliaceae family: *Tilia kiusiana*. The chloroplast DNA (cpDNA) used in the present study included the following three regions: *trnL-trnF* spacer, *trnL* intron and the partial region of the *matK* gene, which encodes a splicing-associated maturase (Neuhaus, and Link, 1987).

MATERIALS AND METHODS

Species sampling The total number of Dipterocarpaceae species included in this study was 79. This includes 42 species from Malaysia, 34 species from Sri Lanka and a single species from each of the following regions: Seychelles, Madagascar and Thailand (Table 1). In addition to sequences obtained in the present study (24 sequences for trnL-trnF spacer and trnL intron regions and 65 sequences for matK), we used data reported in the previous studies (Table 1). For the trnL-trnF spacer and trnL intron regions we used seven sequences obtained by Kamiya et al. (1998), 34 sequences obtained by Gamage et al. (2003) and 14 sequences reported by Kajita et al. (1998). For the matK region, we used 14 sequences from Malaysian species reported by Kajita et al. (1998). The

Table 1. List of species used in this study and the database accession numbers of the DNA sequences

Species	Source	Database Accession No.			
		trnL- $trnF$	trnL	matK	Keterence
Family: Dipterocarpaceae					
Sub family: Dipterocarpoideae					
Tribe: Shoreae					
Dryobalanops aromatica C. F. Gaertn	Frim, Kepong, Malayasia	AB006411	AB006394	AB006377	Kajita et al. (1998)
Dryobalanaops oblongofolia Dyer	Frim, Kepong, Malayasia	AB006412	AB006395	AB006378	Kajita et al. (1998)
Hopea nervosa King	Frim, Kepong, Malayasia	AB006418	AB006401	AB006384	Kajita et al. (1998)
Hopea jucunda Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246524	AB246589	AB246459	Present study, Gamage et al. (2003)
Hopea discolor Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246523	AB246588	AB246458	Present study, Gamage et al. (2003)
Hopea jucunda ssp. modesta DC.	Kanneliya Forest Reserve, Sri Lanka	AB246525	AB246590	AB246460	Present study, Gamage et al. (2003)
Hopea subalata Sym.	Frim Arboretum, Malayasia	AB246520	AB246585	AB246455	Present study, Gamage et al. (2003)
Hopea wightiana Wall.	Frim Arboretum, Malayasia	AB246526	AB246591	AB246461	Present study, Gamage et al. (2003)
Hopea odorata Roxb.	Frim, Kepong, Malayasia	AB006419	AB006402	AB006385	Kajita et al. (1998)
Hopea latifolia Sym.	Frim Arboretum, Malayasia	AB246521	AB246586	AB246456	Present study
Hopea helferi (Dyer) Brandis	Frim Arboretum, Malayasia	AB246522	AB246587	AB246457	Present study
Neobalanocarpus heimii (King) Ashton	Frim, Kepong, Malayasia	AB006417	AB006400	AB006383	Kajita et al. (1998)
Parashorea lucida (Miq.) Kurz	Frim, Kepong, Malayasia	AB006416	AB006399	AB006382	Kajita et al. (1998)
Shorea seminis (de Vriese) Slooten	Frim Arboretum, Malayasia	AB246515	AB246580	AB246450	Present study
Shorea elliptica Burck	Engkabang, Semengoh, Malayasia	AB246509	AB246574	AB246444	Present study
Shorea splendens Ashton	Engkabang, Semengoh, Malayasia	AB246508	AB246573	AB246443	Present study
Shorea pinanga Scheff.	Engkabang, Semengoh, Malayasia	AB246510	AB246575	AB246445	Present study
Shorea acuminata Dyer	Mersing Johor, Malayasia	AB246505	AB246570	AB246440	Present study
Shorea leprosula Miq.	Seremban, Negeri Sembilan, Malayasia	AB246504	AB246569	AB246439	Present study
Shorea xanthophylla Sym.	Frim Arboretum, Malayasia	AB246517	AB246582	AB246452	Present Study, Kamiya et al. (1998)
Shorea bullata Ashton	Engkabang, Semengoh, Malayasia	AB246500	AB246565	AB246435	Present Study, Kamiya et al. (1998)
Shorea curtisii Dyer ex King	Mersing Johor, Malayasia	AB246498	AB246563	AB246433	Present Study, Kamiya et al. (1998)
Shorea macroptera Dyer	Frim, Kepong, Malayasia	AB246503	AB246568	AB246438	Present study
Shorea parvifolia Dyer	Seremban, Negeri Sembilan, Malayasia	AB246502	AB246567	AB246437	Present study, Gamage et al. (2003)
Shorea quadrinervis Sloot.	Kubah National Park, Malayasia	AB246501	AB246566	AB246436	Present study, Kamiya et al. (1998)
Sorea bracteolata Dyer	Frim, Kepong, Malayasia	AB006415	AB006398	AB006381	Kajita et al. (1998)
Shorea ovalis (Korth.)	Frim, Kepong, Malayasia	AB006414	AB006397	AB006380	Kajita et al. (1998)
Shorea macrophylla (de Vriese) Ashton	Frim Arboretum, Malayasia	AB246506	AB246571	AB246441	Present study, Kamiya et al. (1998)
Shorea fallax Meijer	Kubah National Park, Malayasia	AB246499	AB246564	AB246434	Present study, Kamiya et al. (1998)
Shorea richetia Sym.	Kubah National Park, Malayasia	AB246507	AB246572	AB246442	Present study, Gamage et al. (2003)
Shorea laevis Ridl.	Frim Arboretum, Malayasia	AB246514	AB246579	AB246449	Present study
Shorea multiflora (Burck) Sym.	Semengoh Arboretum, Malayasia	AB246516	AB246581	AB246451	Present study
Shorea assamica Dyer	Frim Arboretum, Malayasia	AB246518	AB246583	AB246453	Present study, Gamage et al. (2003)
Shorea congestiflora Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246528	AB246593	AB246463	Present study, Gamage et al. (2003)
Shorea worthingtonii Ashton	Kanneliya Forest Reserve, Sri Lanka	AB246534	AB246599	AB246469	Present study
Shorea pallescens Ashton	Kanneliya Forest Reserve, Sri Lanka	AB246513	AB246578	AB246448	Present study, Gamage et al. (2003)

D. T. GAMAGE et al.

Table 1. Continued

		Database Accession No.			
Species	Source	trnL- $trnF$	trnL	matK	Reference
Shorea disticha (Thw.) Ashton	Gilimale Forest, Sri Lanka	AB246530	AB246595	AB246465	Present study
Shorea megistophylla Ashton	Royal Botanical Garden, Sri Lanka	AB246529	AB246594	AB246464	Present study, Gamage et al. (2003)
Shorea zeylanica (Thw.) Ashton	Royal Botanical Garden, Sri Lanka	AB246535	AB246600	AB246470	Present study, Gamage et al. (2003)
Shorea trapezifolia (Thw.) Ashton	Kanneliya Forest Reserve, Sri Lanka	AB246531	AB246596	AB246466	Present study, Gamage et al. (2003)
Shorea affinis (Thw.) Ashton	Kottawa Arboretum, Sri Lanka	AB246536	AB246601	AB246471	Present study, Gamage et al. (2003)
Shorea cordifolia (Thw.) Ashton	Kanneliya Forest Reserve, Sri Lanka	AB246527	AB246592	AB246462	Present study, Gamage et al. (2003)
Shorea stipularis Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246519	AB246584	AB246454	Present study, Gamage et al. (2003)
Shorea lissophylla Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246512	AB246577	AB246447	Present study
Shorea ovalifolia (Thw.) Ashton	Gilimale Forest, Sri Lanka	AB246532	AB246597	AB246467	Present study
Shorea gardneri (Thw.) Ashton	Bambarabotuwa Forest Reserve, Sri Lanka	AB246533	AB246598	AB246468	Present study
Shorea dyeri Thw.	Bambarabotuwa Forest Reserve, Sri Lanka	AB246511	AB246576	AB246446	Present study
Tribe: Dipterocarpeae					
Anisoptera oblonga Dyer	Frim, Kepong, Malayasia	AB006405	AB006388	AB006371	Kajita et al. (1998)
Anisoptera laevis Dyer	Frim, Kepong, Malayasia	AB006404	AB006387	AB006370	Kajita et al. (1998)
Cotylelobium malayanum V. Sl.	Frim Arboretum, Malayasia	AB246479	AB246544	AB246414	Present study
Cotylelobium scabriusculum (Thw.) Brandis	Kottawa Forest, Sri Lanka	AB246480	AB246545	AB246415	Present study, Gamage et al. (2003)
Dipterocarpus alatus A. DC.	Samui Island, Thailand	AB246538	AB246603	AB246473	Present study
Dipterocarpus cornutus Dyer	Frim Arboretum, Malayasia	AB246537	AB246602	AB246472	Present study
Dipterocarpus baudii Korth	Frim, Kepong, Malayasia	AB006410	AB006393	AB006376	Kajita et al. (1998)
Dipterocarpus kerrii King	Frim, Kepong, Malayasia	AB006409	AB006392	AB006375	Kajita et al. (1998)
Dipterocarpus glandulosus Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246542	AB246607	AB246477	Present study, Gamage et al. (2003)
Dipterocarpus hispidus Thw.	Wilpita Forest, Sri Lanka	AB246541	AB246606	AB246476	Present study, Gamage et al. (2003)
Dipterocarpus zeylanicus Thw.	Diyadawa Forest, Sri Lanka	AB246539	AB246604	AB246474	Present study, Gamage et al. (2003)
Dipterocarpus insignis Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246540	AB246605	AB246475	Present study, Gamage et al. (2003)
Stemonoporus acuminatus (Thw.) Beddome	Kanneliya Forest Reserve, Sri Lanka	AB246487	AB246552	AB246422	Present study, Gamage et al. (2003)
Stemonoporus lancifolius (Thw.) Ashton	Kanneliya Forest Reserve, Sri Lanka	AB246495	AB246560	AB246430	Present study
Stemonoporus kanneliyensis Kosterm.	Kanneliya Forest Reserve, Sri Lanka	AB246494	AB246559	AB246429	Present study, Gamage et al. (2003)
Stemonoporus canaliculatus Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246490	AB246555	AB246425	Present study, Gamage et al. (2003)
Stemonoporus bullatus Kosterm.	Kanneliya Forest Reserve, Sri Lanka	AB246491	AB246556	AB246426	Present study, Gamage et al. (2003)
Stemonoporus reticulatus Thw.	Kanneliya Forest Reserve, Sri Lanka	AB246492	AB246557	AB246427	Present study, Gamage et al. (2003)
Stemonoporus scalarinervis Kosterm.	Gilimale Forest, Sri Lanka	AB246489	AB246554	AB246424	Present study, Gamage et al. (2003)
Stemonoporus wightii Thw.	Gilimale Forest, Sri Lanka	AB246493	AB246558	AB246428	Present study
Stemonoporus gilimalensis Kosterm.	Gilimale Forest, Sri Lanka	AB246488	AB246553	AB246423	Present study
Upuna borneensis Sym.	Frim, Kepong, Malayasia	AB006408	AB006391	AB006374	Kajita et al. (1998)
Vatica coriacea Ashton	Kubah National Park, Malayasia	AB246483	AB246548	AB246418	Present study
Vatica micrantha V. Sl.	Kubah National Park, Malayasia	AB246484	AB246549	AB246419	Present study, Gamage et al. (2003)
Vatica affinis Thw.	Kottawa, Sri Lanka	AB246486	AB246551	AB246421	Present study, Gamage et al. (2003)
Vatica pauciflora (Korth.) BI.	Frim Arboretum, Malayasia	AB246482	AB246547	AB246417	Present study, Gamage et al. (2003)
Vatica odorata Roxb.	Frim, Kepong, Malayasia	AB006419	AB006402	AB006385	Kajita et al. (1998)
Vatica bella V. Sl.	Frim Arboretum, Malayasia	AB246481	AB246546	AB246416	Present study, Gamage et al. (2003)
Vatica chinensis L.	Gilimale. Sri Lanka	AB246485	AB246550	AB246420	Present study, Gamage et al. (2003)
Vateria copallifera (Retzius) Alston	Kanneliya Forest Reserve, Sri Lanka	AB246496	AB246561	AB246431	Present study, Gamage et al. (2003)
Vateriopsis seychellarum Dver	Sevchelles	AB246497	AB246562	AB246432	Present study, Gamage et al. (2003)
Sub-family: Monotoideae	U				(2000)
Monotes madagascariensis Humb.	Fenetrede l'Isalo, Madagascar	AB246543	AB246608	AB246478	Present study, Gamage et al. (2003)
Family: Tiliaceae					
Tilia kiusiana Makino et Shirasawa	Kyushu University, Japan	AB006420	AB006403	AB006386	Kajita et al. (1998)

Sequence (5'-3')	Usage	Reference
GGTTCAAGTCCCTCTATCCC	PCR and sequencing	Taberlet et al. (1991)
ATTTGAACTGGTGACACGAG	PCR and sequencing	Taberlet et al. (1991)
CGAAATCGGTAGACGCTACG	PCR and sequencing	Taberlet et al. (1991)
GGGGATAGAGGGACTTGAAC	PCR and sequencing	Taberlet et al. (1991)
CTATATCCACTTATCTTTCAGGAGT	PCR and sequencing	Ooi et al. (1995)
CTGCATATACGCCCAAATCGGTCAA	PCR and sequencing	Ooi et al. (1995)
GAAATGCGGGTTCGACA	PCR and sequencing	Present study
GGACAATGATCCAATCAAGGC	PCR and sequencing	Present study
TCAGTTTACTGATTGTAAAACG	PCR and sequencing	Present study
TTTGGACAATGATCCAATCAAG	PCR and sequencing	Present study
TCCAGATCGGCTTACTAATG	Sequencing	Kajita et al. (1998)
GATGGATGGGATGAGGTATTAGT	Sequencing	Kajita et al. (1998)
AATGGATTCGTATTCACA	Sequencing	Present study
GTATGTGAATACGAATCCAT	Sequencing	Present study
	Sequence (5'-3') GTTCAAGTCCCTCTATCCC TTTGAACTGGTGACACGAG GGAAATCGGTAGACGCTACG GGGATAGAGGGACTTGAAC TATATCCACTTATCTTTCAGGAGT TGCATATACGCCCAAATCGGTCAA GAAATGCGGGTTCGACA GACAATGATGCAATCAAGGC 'CAGTTTACTGATTGTAAAACG TTGGACAATGATCCAATCAAGGC CCAGATCGGCTTACTAATG GATGGATGGGATG	Sequence (5'-3')UsageGGTTCAAGTCCCTCTATCCCPCR and sequencingGTTGAACTGGTGACACGAGPCR and sequencingCGAAATCGGTAGACGCTACGPCR and sequencingCGGAAATCGGTAGACGCTACGPCR and sequencingCGGAAATCGGTAGACGCTACGPCR and sequencingCGGAAATCGGTAGACGCTACGPCR and sequencingCTATATCCACTTATCTTTCAGGAGTPCR and sequencingCTGCATATACGCCCAAATCGGTCAAPCR and sequencingCAAATGCGGGTTCGACAPCR and sequencingCACAATGATCCAATCAAGGCPCR and sequencingCAGTTTACTGATTGTAAAACGPCR and sequencingCCAGATCGGCTTACTAATGSequencingCCAGATCGGCTTACTAATGSequencingATGGATGGGATGAGGTATTAGTSequencingATGGATTCGTATTCACASequencingCTATGTGAAAACGAATCCAATCAAGSequencing

Table 2. Primers used in the present study

remaining matK sequences (65) were obtained in the present study. In addition, we included sequences of the trnL-trnF spacer, trnL intron and the matK gene of *Tilia* kiusiana, which belongs to the family Tiliaceae.

DNA isolation, PCR and sequencing Total DNA was extracted as described by Gamage et al. (2003). The intergenic spacer region between trnL and trnF genes, *trnL* intron region and a partial region of the *matK* gene were amplified by polymerase chain reaction (PCR). The primers designed by Taberlet et al. (1991) were used to amplify the *trnL-trnF* spacer and *trnL* intron regions. The primers designed by Ooi et al. (1995) were used to amplify the partial matK gene region. Since for most samples the *matK* gene region could not be amplified, several new primers for both PCR and sequencing were designed. The primers used in this study are listed in Table 2. Amplification was carried out after denaturing the DNA at 94°C for 3 minutes followed by 30 cycles of 1 minute at 94°C, 1 minute at 52–55°C for annealing, 1.3 minutes at 72°C, and ending with 7 minutes at 72°C for extension.

PCR products were purified using MiniElute PCR Purification QIAGEN Kit according to the manufacturer's instructions. Sequencing reactions were carried out using the BigDyeTM Terminator v.3.1 Cycle Sequencing Ready Reaction Kit (Applied Biosystems) following the manufacturer's instructions. Purified PCR products were directly sequenced using the ABI Prism 3100 Genetic Analyzer (Applied Biosystems). The sequences were determined in both directions. For the *trnL-trnF* spacer and *trnL* intron regions sequencing primers were the same as those used for PCR, while sequencing primers designed by Kajita et al. (1998) and newly designed PCR

and internal primers were used for sequencing of the *matK* region (Table 2). Nucleotide sequence data obtained in this study and the sequences used by Kamiya et al. (1998) are diposited in the DDBJ/EMBL/GenBank databases under accession numbers AB246414 through AB246478, AB246479 through AB246543 and AB246544 through AB246608 for the *matK*, *trnL-trnF*, *and trnL* intron regions respectively.

Data analyses Sequences for the trnL-trnF spacer, trnL intron and matK regions were aligned individually and as combined data set using the ClustalX program (Thompson et al., 1997). The aligned sequences were corrected manually using the BioEdit program (Hall, 1999).

Kimura's two-parameter distance (Kimura, 1980) was used to calculate the genetic distances for all pairs of sequences. Neighbor-joining (NJ) trees (Saitou, and Nei, 1987) for both individual and combined data sets were constructed excluding and including alignment gaps using the ClustalX program (Thompson et al., 1997). Pair-wise deletion option was used when gaps were included in the distance calculations. Phylogenetic tree using Maximum Likelihood (ML) method (Felsenstein, 1981) was obtained based on the combined data using the SEQBOOT, DNAML and CONSENSE programs from the PHYLIP v. 3.6 package (Felsenstein, 2004). In this analysis, empirical nucleotide frequencies were used and the transition/transversion ratio was set to 0.5 as estimated from the combined data set. The statistical support for the nodes of the trees was determined using bootstrap (BT) method (Felsenstein, 1985) based on 1000 replicates.



Fig. 1. Neighbor joining tree constructed using the combined data set for the trnL-trnF spacer, trnL intron and matK regions, based on Kimura's two parameter distance (Kimura, 1980). Bootstrap values (BT) in percent from 1000 replicates are indicated above the nodes. The BT values, which were < 50% are not shown. The tree is unrooted and branch lengths are proportional to the scale given in nucleotide substitution per site.

RESULTS

Characteristics of trnL-trnF spacer, trnL intron and *matK* sequences In the present study, we determined sequences of the trnL-trnF spacer and trnL intron regions for additional 24 species, which were not included in our previous study (Gamage et al. 2003). For the trnL-trnF spacer region the total number of sites, including gaps was 408 after alignment of which 101 sites were variable and 49 were parsimony informative. For the trnL intron region the total number of sites after alignment, including gaps was 560 of which 165 sites were variable and 39 were parsimony informative. There were 14 indels ranging from 1 bp to 89 bp in the *trnL-trnF* spacer while 30 indels ranging from 1 bp to 101 bp were found in the *trnL* intron. A long indel of 89 bp was found in the *trnL*-trnF spacer region of Hopea latifolia and H. helferi. The longest indel, 101 bp was found in the trnL intron of all Stemonoporus species. Sri Lankan endemic species belonging to the Doona genus established by Kostermans (1984) and Maury (1978) had a common 33 bp indel in the trnL intron. All species from the genus Vat*ica* had one 6 bp indel in both the *trnL-trnF* spacer and the trnL intron regions. Monotes madagascariensis had two indels of 1 bp and 4 bp in the trnL-trnF spacer and one indel of 7 bp in the trnL intron. Tilia kiusiana had eight indels (<4bp) in the trnL-trnF spacer and three indels (two of 1 bp and one of 38 bp) in the trnL intron.

In the present study, we determined 65 sequences of the partial matK gene region. The aligned matrix of the matK region comprised 972 bp. There were 235 polymorphic sites, of which 106 sites were parsimony informative. One 6 bp long indel was found in the *T. kiusiana* sequence.

Phylogenetic analysis Topologies of the NJ trees obtained separately for the *trnL*-trnF spacer, *trnL* intron, and the partial *matK* gene region were generally congruent, with small differences in the resolution of some genera such as Cotylelobium, Dipterocarpus, Hopea and Vateriopsis (data not shown). The topology of the NJ tree based on the *matK* sequence was identical with that of the tree based on the combined data set including all three regions used in the present study (data not shown). There were no considerable topological differences between the trees constructed including and excluding alignment gaps. The topology and BT support of the ML tree based on the combined data set were very similar to those of the corresponding NJ tree (data not shown). Therefore, only the NJ tree constructed using the combined data set including alignment gaps is presented (Fig. 1).

Monotes madagascariensis was grouped together with Tilia kiusiana. Next to this group was Vateriopsis seychellarum, which occupied a single branch sister to the clade containing species from the Dipterocarpeae tribe. The genera: Stemonoporus, Anisoptera, Vateria, Upuna, Cotylelobium and Vatica formed distinct clade (designated as A on Fig. 1) supported by high bootstrap (BT) probability (99%). It contained two clades: the Stemonoporus clade, and the clade with the following genera: Anisoptera, Vateria, Upuna, Cotylelobium and Vatica. Within the latter clade Anisoptera, Cotylelobium and Vatica were monophyletic, while Upuna and Vateria formed sister clades. Except for the Upuna, Vateria and Vatica clades, other generic clades had high BT support (80%– 99%).

Most of the remaining genera included in our study formed separate clades on our tree. Here, the genera *Dipterocarpus* and *Dryobalanops* formed two separate monophyletic clades each with 100% BT support. *Dipterocarpus* clade was divided into two groups, one (BT = 77%) with Malaysian species (*D. kerrii*, *D. baudii* and *D. cornutus*) and the other (BT = 71%) with three Sri Lankan species (*D. zeylanicus*, *D. insignis*, and *D. glandulosus*) and *D. alatus* from Thailand.

The remaining genera formed two main clades (designated as B and C on Fig. 1). Clade B contained species belonging to the genus *Richetia* (BT = 100%) (Maury, 1978) or Yellow Meranti (Symington, 1943) wood group, Shorea (Maury, 1978) or Balau (Symington, 1943) wood group, Parashorea (BT = 65%), Rubroshorea (BT = 81%) (Maury, 1978) or Red Meranti wood group (Symington, 1943). The clade C contained the following genera: *Doona* (BT = 99%) (Kostermans, 1984; Kostermans, 1992; Maury, 1978), Anthoshorea (BT = 99%) (Maury, 1978) or White Meranti wood group (Symington, 1943), Neobalanocarpus heimii (BT = 97%) and Hopea (BT = 97%). The clades B and C had 93% BT support. Species belonging to the genera Richetia and Shorea formed separate clades. Parashorea and Rubroshorea were grouped separately with the former genus as a sister clade. Richetia clade was sister to the Shorea clade.

DISCUSSION

Phylogeny reconstruction We obtained two NJ trees (with and without alignment gaps) for the combined data set including trnL-trnF spacer, trnL intron, and the partial region of the matK gene. Except for the differences in the bootstrap (BT) support for some nodes, the topologies of these trees were similar. Thus, we think that alignment gaps had little effect on the topology of our phylogenetic trees. Furthermore, topologies of our NJ and ML trees were nearly identical. Hence, only the NJ tree based on combined data set including gaps is discussed.

Generic relationships The generic relationships revealed by our NJ tree are mostly in agreement with previous molecular phylogenies based on cpDNA (Dayanandan et al., 1999; Gamage et al., 2003; Kajita et al., 1998; Kamiya et al., 1998). However, our current results provide new evidence regarding the relationships of additional genera such as *Monotes*, *Vateriopsis* and *Stemonoporus*, which were not included or discussed well (except *Monotes*) in most previous studies (Dayanandan et al., 1999; Kajita et al., 1998; Kamiya et al., 1998). Our study also gives new information about the validity of the previous classifications of the genus *Shorea*.

To date, the genus Monotes was included only in three previous studies of Dipterocarpaceae phylogeny (Dayanandan et al., 1999; Gamage et al., 2003; Morton et al., 1999). Moreover, no effort employing molecular data was made to directly test its placement in the family Tiliaceae suggested by Heim (1892) based on flowers and fruit features. Morphological similarity of Monotes to Tiliaceae was also suggested by Kostermans (1985). Other classifications however, placed it in a separate family Monotaceae, which includes two sub-families Monotoideae and Pakaraimoideae (Kostermans, 1989). Based on the *rbcL* sequences, Dayanandan et al. (1999) suggested that Monotes is more related to the Asian Dipterocarpaceae than to Tiliaceae. Our present results showed that Monotes madagascariensis was placed together with Tilia kiusiana. However, the M. madagascariensis branch was much shorter (0.0243) than the branch with T. kiusiana (0.1159). Furthermore, the internal branch of the clade containing these two species was relatively short (0.0080). This result supports suggestion by Dayanandan et al. (1999) to place the genus Monotes within the Dipterocarpaceae family. Furthermore, it also indicates that *M. madagascariensis* appears to be better candidate for outgroup species than T. kiusiana, which was used for this purpose by Kajita et al. (1998).

The topology of the present phylogenetic tree, was to a certain extent, consistent with the current division of the sub-family Dipterocarpoideae into two tribes: Dipterocarpeae (n = 11) and Shoreae (n = 7) (Ashton, 1982; Brandis, 1895; Jong, and Kaur, 1979; Maury-Lechon, 1979). The two tribes formed two monophyletic clades (BT = 98%) on our tree except for the genus *Dipterocarpus* from the Dipterocarpeae tribe, which was placed as a sister clade to species from the Shoreae tribe.

Similar to result reported by Gamage et al. (2003) monotypic species Vateriopsis seychellarum endemic to Seychelles Island was placed on a separate branch sister to the Dipterocarpeae clade (Fig. 1). On our tree, the relationship of this species with other species of the tribe Dipterocarpeae, which have the same chromosome number (n = 11) is relatively well supported (BT = 86%). The origin of this species is still unclear and the possibilities of both plate tectonic movements and the human transportation should be considered (Kostermans, 1992). Embryological evidence suggested that it is related to Dipterocarpus, Hopea, Shorea and Vateria (Oginuma et al., 1999). However, it more resembles *Dipterocarpus* than the other three genera in having the micropyle formed by both the inner and outer integument and a conspicuously enlarged chalaza (basal part of the ovule opposite the micropyle, where integument and nucellus are joined) with ample vascular tissues (Oginuma et al., 1999). The placement of *Vateriopsis seychellarum* on our tree did not support its relationship with *Dipterocarpus*, which occupied a separate clade, sister to the clade containing species from the Shoreae tribe. Based on our results it appears that it rather represents a relatively diverged member of the Dipterocarpeae or Shoreae tribe.

The genus Stemonoporus, which is endemic to Sri Lanka formed a distinct and well supported monophyletic clade (BT = 98%). This is in agreement with the phylogenetic analyses based on *rbcL* data and noncoding cpDNA (Dayanandan et al., 1999; Gamage et al., 2003). Based on comparative morphology Stemonoporus was considered as one of the most archaic genera of the Asian sub-family Dipterocarpoideae (Ashton, and Gunatilleke, 1987). We found a long 101 bp indel in the *trnL* intron in all Stemonoporus species included in the present study. Divergent status of *Stemonoporus* revealed in the present and other studies is also consistent with its unique morphological features such as peculiar anthers with apical dehiscence and apical leaf traces, which separate from the central vascular cylinder well before the node (Ashton, 1982; Kostermans, 1992).

Our present results showed that (except for Upuna and Vateria) Anisoptera, Vatica and Cotylelobium clades are monophyletic, although only the Cotylelobium clade had high BT support (93%). Vatica also showed monophyly in the study by Dayanandan et al. (1999). However, its relationships with Anisoptera and Cotylelobium were not elucidated. On our tree, Upuna and Vateria were grouped together but with low BT support. On the other hand, the rbcL analysis placed Upuna and Vateria on two separate branches sister to Stemonoporus (Dayanandan et al., 1999). Kostermans (1992) suggested that Vateria is closely related with Vatica and that there is no consensus whether these two genera should be fused or kept separate. Further studies are necessary to elucidate the relationship of these two genera.

On our NJ tree, the genus *Dipterocarpus* formed a distinct, highly supported monophyletic clade (BT = 100%, Fig. 1). Similar results were reported in the previous molecular phylogenies (Gamage et al., 2003; Kajita et al., 1998; Kamiya et al., 1998). Morphological evidence also supports highly divergent character of this genus. *Dipterocarpus* has many unique characters, including the winged free calyx tube and large flowers (Dayanandan et al., 1999). Some studies suggested that *Dipterocarpus* might represent the basal clade of Dipterocarpoideae subfamily (Meijer, 1979). On the other hand, others placed *Dipterocarpus* (together with other members of the Dipterocarpeae tribe) as a sister to the group with species of the tribe Shoreae (Maury, 1978). Our present results also indicate that the genus *Dipterocarpus* was among the most diverged genera of the sub-family Dipterocarpoideae. Finally, Sri Lankan *Dipterocarpus* species (*D. glandulosus*, *D. hispidus*, *D. insignis* and *D. zeylanicus*) formed a separate clade but they were not much diverged from the other species of the *Dipterocarpus* clade. Thus, our results suggest early divergence of this genus from other species of the Dipterocarpoideae and independent evolution of Sri Lankan species.

Similar to *Dipterocarpus*, the genus *Dryobalanops* also formed a distinct, highly supported monophyletic clade on our tree (Fig. 1, BT = 100%). Ashton (1979) placed *Dryobalanops* in the tribe Shoreae due to the presence of connate petals. Such placement was also suggested by the presence of solitary vessels (Gotwald, and Parameswaran, 1966) and the chromosome number (n = 7) (Jong, and Kaur, 1979). However, Maury-Lechon (1979) placed it in the tribe Dipterocarpeae based on the presence of valvate fruit sepals. Our results showed that *Dryobalanops* was placed as a sister clade to the cluster containing species from the Shoreae tribe, which supports classification proposed by Ashton (1979).

Relationships of Shorea, Hopea, Neobalanocarpus and Parashorea The topology of our tree lends some support to the classification of Shorea species proposed by Maury (1978) and Maury-Lechon (1979). On our tree, the genera created by this author (Richetia, Shorea, Rubroshorea, Doona and Anthoshorea) are resolved as separate groups, although the clade containing Shorea members had weak BT support (<50%). Some of these genera (Richetia, Shorea and Rubroshorea) also formed separate groups on the tree reported by Kamiya et al. (2005) although their study did not include Doona and Pentacme species. Our present study also did not include species from the genus *Pentacme* recognized by Maury (1978) and Maury-Lechon (1979). Therefore, it is important to include them in future phylogenies for obtaining further support for the classification of Shorea species proposed by this author.

The placement of *Shorea* species on our tree is also in agreement with classification proposed by Symington (1943). White Meranti, Red Meranti, Balau and Yellow Meranti were all monophyletic. Similar result, except monophyly of the Red Meranti, was reported by Gamage et al. (2003) and Kamiya et al. (1998). On our tree, White Meranti, Red Meranti, and Yellow Meranti had high BT support (>81%). However, the Yellow Meranti-Balau clade was sister to the clade with *Parashorea* and Red Meranti. Thus, the topology of our tree is not consistent with that of the tree obtained by Kamiya et al. (2005), where *Parashorea* was placed on a long separate branch sister to Yellow Meranti, Balau and Red Meranti. On the other hand, the placement of *Parashorea* (within the clade containing Balau, Red Meranti and White Meranti) revealed in our present study is similar to that reported in the previous cpDNA based phylogenies (Gamage et al., 2003; Kamiya et al., 1998). Actually, the wood groups recognized by Symington (1943), well correspond with the generic classification proposed by Maury (1978) and Maury-Lechon (1979). That is, Yellow Meranti with *Richetia*, Balau with *Shorea*, Red Meranti with *Rubroshorea* and White Meranti with *Anthoshorea*.

Taking into account many distinctive morphological differences between Shorea and Doona species, several studies suggested that *Doona* should be regarded as a separate genus (Kostermans, 1984; Kostermans, 1992; Maury, 1978; Maury-Lechon, 1979). In our present study, species placed by these authors in the genus Doona (S. megistophyla, S. ovalifolia, S. worthingtonii, S. gardneri, S. trapezifolia, S. zeylanica, S. disticha, S. cordifolia, S. congestiflora and S. affinis) formed particularly distinct, monophyletic clade with 99% BT support. Based on morphology these species were placed by Ashton (1972), Ashton (1977) and Ashton (1982) in a separate section Doona. The common 33 bp indel in the trnL intron present in all Doona species provides further evidence for distinct character of this group. Our present study also resolved the position of additional two Doona species (S. disticha and S. ovalifolia), which were not included in our previous study (Gamage et al., 2003). Therefore, we could determine phylogenetic position of almost all the Doona species present in Sri Lanka.

Parameswaran and Gotwald (1979) reported that the genus Neobalanocarpus has close affinity with Doona based on wood anatomy. Floral characters such as diurnal anthesis and stamen structure of Neobalanocarpus also show similarity to Doona (Dayanandan et al., 1999). However, on our tree Doona had the sister relationship to the Neobalanocarpus heimii branch, which in turn was sister to the Hopea clade. This agrees with results of other cpDNA phylogenies (Gamage et al., 2003) but is incongruent with phylogeny based on nuclear (n) DNA, which placed Neobalanocarpus together with Anthoshorea species in the most basal and first diverged clade sister to clades containing Shorea, Parashorea and Hopea (Kamiya et al., 2005). The different placement of Neobalanocarpus in cpDNA and nDNA based phylogenies together with the morphological characters shared by Neobalanocarpus, Anthoshorea and Hopea, and the irregular behavior of Neobalanocarpus during meiosis (Jong, and Lethbridge, 1967) lead Kamiya et al. (2005) to suggest that it may be a hybrid between Anthoshorea and Hopea. Our results also showed that Neobalanocarpus has an intermediate position between Hopea and Anthoshorea (BT = 97%). If this placement is associated with the hybrid nature of Neobalanocarpus our present result would also imply the occurrence of recombination in the

cpDNA. This is surprising because it is believed that due to its uniparental inheritance in most plants cpDNA does not undergo recombination (Chiu and Sears, 1985). Therefore, our result suggests that cpDNA in some Dipterocarpaceae species is inherited biparentally and undergoes recombination. Further investigation regarding this mater is necessary.

An unresolved feature in the previous classifications of the Shoreae tribe was that the well recognized genus Hopea was placed within the clade containing other species of that tribe. Our results showed that Hopea group was monophyletic within the clade containing Anthoshorea and Neobalanocarpus and had high BT support (97%). The topologies of the *PgiC* (Kamiva et al., 2005) and rbcL trees (Dayanandan et al., 1999) also showed the monophyly of Hopea and placed it within the clade containing other species of the Shoreae tribe. The floral morphology of the genera Hopea and Anthoshorea are similar, both having an urceolate corolla and stamens with an acicular connective appendage (Dayanandan et al., 1999). There are also some unique morphological characters shared by Hopea, some Shorea species and Neobalanocarpus (Kamiya et al., 2005). Therefore, it is possible that these genera have yet not reached the generic level of divergence at molecular level, even though they have already evolved some different morphological characters. Further evaluation using morphological and molecular data is important for detailed classification of these genera.

Relationships of Sri Lankan species There are about 58 species of Dipterocarpaceae in Sri Lanka (Kostermans, 1992). They belong to the genera: Dipterocarpus, Shorea, Doona, Hopea, Stemonoporus, Cotylelobium, Vatica, and Vateria. Ninety eight percent of the species are endemic. Our present phylogeny revealed the monophyly of Sri Lankan endemic genus Stemonoporus and Doona species while other species formed separate clades. However, the present phylogeny did not reveal much divergence between Sri Lankan and other Dipterocarpideae species. The isolated position of Sri Lankan species on our tree may be due to their independent evolution caused by the geographic isolation. The placement of Sri Lankan Shorea stipularis, which belongs to the Anthoshorea section (Ashton, 1980; Ashton, 1982) or genus (Maury, 1978) with other Anthoshorea species (S. bracteolata and S. assamica) from Malaysia is in agreement with such taxonomical grouping. Geographical distribution of S. stipularis in Sri Lanka and its morphological similarity to Malaysian Shorea species suggest that Dipterocarpaceae must have already diverged to generic or infrageneric sections before they entered the Laurasian plate from the Deccan plate according to the Gondwanan origin of Asian Dipterocarpaceae (Dayanandan et al., 1999). The other Sri Lankan Shorea species

(S. lissophylla, S. dyeri and S. pallescens), which belong to section (Ashton, 1982) or genus (Maury, 1978) Shorea were monophyletic and had close relationship with other species from this group. To obtain more refined phylogeny and further insights into the evolutionary history of Sri Lankan species, additional sequence data are necessary. They should also include species from India. Sri Lanka was intermittently connected to mainland India and this could have enabled biotic interchange with southern India during the Pleistocene ice ages (Bossuyt et al., 2004). Thus, Sri Lankan and Indian Dipterocarpaceae species may be closely related. The most likely possibility is that Dipterocarpaceae spread to Sri Lanka through India.

We would like to thank H. Ishiyama from Kyushu University, Japan and A. Yoshida from the Research Institute of Evolutionary Biology, Japan and two anonymous reviewers for help and comments in the course of this work, K. Kamiya also from Kyushu University, Japan for providing unpublished sequence data for some Shorea species, M. Jayaweera from the Plant Genetic Resources Center, Sri Lanka for identifying Dipterocarpaceae species collected from Sri Lanka, the Officers of the Forest Department and Forest Conservation Department, Sri Lanka for permission to collect samples. This work was financially supported by the grant (No.1535 SRI (SF)) to the Department of Botany, University of Ruhuna, Sri Lanka from the Asian Development Bank and the grants 13490022 and 13575002 to AES and 072898 to TY from the Ministry of Education, Culture, Sports, Science and Technology, Japan and the Sasakawa Scientific Research grant 16-268 to TGD from the Japan Science Society.

REFERENCES

- Ashton, P. S., (1972) Precursor to a taxonomic revision of Ceylon Dipterocarpaceae. Blumea 20, 357–366.
- Ashton, P. S., (1977) Dipterocarpaceae. Revised handbook flora of Ceylon. 2, 166–196.
- Ashton, P. S., (1979) Phylogenetic speculations on Dipterocarpaceae. Histoire Natur. Paris N. S. (B) 26, 145–149.
- Ashton, P. S., (1980) Dipterocarpaceae. In: Revised Handbook to the Flora of Ceylon I (eds.: M. D. Dassanayake, and F. R. Fosberg), pp. 364–423. New Delhi.
- Ashton, P. S., (1982) Dipterocarpaceae. Flora Malesiana 9, 237–552.
- Ashton, P. S., and Gunatilleke, C. V. S., (1987) New light on the plant geography of Ceylon. I Historical plant geography, J. Biogeog. 14, 249–285.
- Bossuyt, F., Meegaskumbura, M., Beenaerts, N., Gower, D. J., Pethiyagoda, R., Roelants, K., Mannaert, A., Wilkinson, M., Bahir, M. M., Arachchi, K. M., Ng, P. K. L., Schneider, C. J., Oommen, O. V., and Milinkovitch, M. C., (2004) Local Endemism Within the Western Ghats-Sri Lanka Biodiversity Hotspot. Science **306**, 479–480.
- Brandis, D., (1895) An enumeration of the Dipterocarpaceae. J. Linnean Soc. **31**, 1–148.
- Chiu, W. L., and Sears, B. B., (1985) Recombination between chloroplast DNAs does not occur in sexual crosses of *Oenothera*. Mol. Gen. Genet. 198, 525–528.
- Dayanandan, S., Ashton, P. S., M., W., and Primack, R. B., (1999) Phylogeny of the tropical tree family Dipterocarpaceae based on nucleotide sequences of the chloroplast

rbcL gene. Am. J. Bot. 86, 1182–1190.

- Felsenstein, J., (1981) Evolutionary trees from DNA sequences: A maximum likelihood approach. J. Mol. Evol. 17, 368– 376.
- Felsenstein, J., (1985) Confidence limits on phylogenies: an approach using the bootstrap. Evolution **39**, 783–791.
- Felsenstein, J., (2004) PHYLIP (Phylogeny Inference Package) version 3.6. Distributed by the author. Department of Genome Science, University of Washington, Seattle.
- Gamage, D. T., de Silva, M., Yoshida, A., Szmidt, A. E., and Yamazaki, T., (2003) Molecular phylogeny of Sri Lankan Dipterocarpaceae in relation to other Asian Dipterocarpaceae based on chloroplast DNA sequences. Tropics 13, 79–87.
- Gilg, E., (1925) Dipterocarpaceae in Engler und Prantl. Natürliche Pflanzenfamilien **21**, 237–269.
- Gotwald, H., and Parameswaran, N., (1966) Das sekundäre Xylem der Familie Dipterocarpaceae, Anatomische Untersuchungen zur Taxonomie und Phylogenie. Bot. Jahrb. Syst. 85, 410–508.
- Hall, T. A., (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/ NT. Nucleic Acids. Symp. 41, 95–98.
- Heim, F., (1892) Recherches sur les Diptérocarpacées. Ph. D. dissertation. à la Faculte des Sciences de Paris. pp. 1–186.
- Jong, K., and Kaur, A., (1979) A cytotaxonomic view of Dipterocarpaceae with some comments on polyploidy and apomixis.
 In: Diptérocarpacées: Taxonomie-Phylogénie-Ecologie (ed.: G. Maury-Lechon), pp. 41–49. Mémoires du Muséum National d 'Histoire Naturelle: Série B, Botanique, Editions du Muséum, Paris.
- Jong, K., and Lethbridge, A., (1967) Cytological studies in the Dipterocarpaceae. I. Chromosome numbers of certain Malaysian genera. Notes from the Royal Garden Edinburgh 27, 175–184.
- Kajita, T., Kamiya, K., Nakamura, K., Tachida, H., Wickneswari, R., Tsumura, Y., Yoshimaru, H., and Yamazaki, T., (1998) Molecular Phylogeny of Dipterocarpaceae in Southeast Asia Based on Nucleotide Sequences of *matK*, *trnL* intron and *trnL-trnF* Intergenic Spacer Region in Chloroplast DNA. Mol. Phyl. Evol. **10**, 202–209.
- Kamiya, K., Harada, K., Ogino, K., Kajita, T., Yamazaki, T., Lee, H. S., and Ashton, P. S., (1998) Molecular Phylogeny of Dipterocarp Species Using Nucleotide Sequences of Two Non-coding Regions in Chloroplast DNA. Tropics 7, 195– 207.
- Kamiya, K., Harada, K., Tachida, H., and Ashton, P. S., (2005) Phylogeny of *PgiC* gene in *Shorea* and its closely related genera (Dipterocarpaceae), the dominant trees in Southeast Asian tropical rain forests. Am. J. Bot. **92**, 775–788.
- Kimura, M., (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J. Mol. Evol. 16, 111–120.
- Kostermans, A. J. G. H., (1978) Pakaraimaea Dipterocarpaceae Maguire and Ashton belongs to Tiliaceae and not to Dipterocarpaceae. Taxon 27, 357–359.
- Kostermans, A. J. G. H., (1982) The genus Hopea (Dipterocarpaceae) in Ceylon, Sri Lanka. Ceylon J. Biol. Sci. 15, 41– 49.
- Kostermans, A. J. G. H., (1984) Monograph of the genus Doona Thw. (Dipterocarpaceae). Botanische Jahrbücher. Systematik Pflazengeschichte und Pflanzengeographie 104, 425– 454.
- Kostermans, A. J. G. H., (1985) Family status of the Monotoideae Gilg. and the Pakaraimoideae Ashton, Maguire and

de Zeeuw (Dipterocarpaceae). Taxon 34, 426-435.

- Kostermans, A. J. G. H., (1989) Monotaceae, a new family allied to Tiliaceae. Taxon **38**, 123–124.
- Kostermans, A. J. G. H., (1992) A handbook of the Dipterocarpaceae of Sri Lanka. The Wildlife Heritage Trust of Sri Lanka, Colombo, Sri Lanka, pp. 243.
- Maury, G., (1978) Dipterocarpacees: du fruit a la plantule. Ph. D. thesis. Universite Paul Sabatier, Toulouse, Paris. pp. 3 vols., IA: 243p., IB: 432p., II: 344p.
- Maury-Lechon, G., (1979) Conséquences taxonomiques de l'étude des caractères des fruits/ germinations, embryons et plantules des Diptérocarpacées. In: Diptérocarpacées: Taxonomie-Phylogénie-Ecologie (ed.: G. Maury-Lechon), pp. 81– 106. Mémoires du Muséum National d'Histoire Naturelle: First International Round Table on Dipterocarpaceae, Série B, Botanique 26, Editions du Muséum, Paris.
- Maury-Lechon, G., and Curtet, L., (1998) Biogeography and Evolutionary Systematics of Dipterocarpaceae. In: A Review of Dipterocarps: Taxonomy, Ecology and Silviculture (eds.: S. Appanah, and J. M. Turnbull), pp. 5–44. Center for International Forest Research, Bogor, Indonesia.
- Meijer, W., (1979) Taxonomic studies in the genus Dipterocarpus. In: Diptérocarpacées: Taxonomie-Phylogénie-Ecologie (ed.: G. Maury-Lechon), pp. 50-56. Mémoires du Muséum National d'Histoire Naturelle: Série B, Botanique 26, Editions du Muséum, Paris.
- Meijer, W., and Wood, G. H. S., (1964) Dipterocarps of Sabah (North Borneo). Sabah forest record No. 5, Forest Department, Sandakan, Art Printing Works, Kuala Lumpur, Malaysia, pp. 1–344.
- Meijer, W., and Wood, G. H. S., (1976) Keys to Dipterocarps on Sabah. Biotrop, Bogor, Indonesia.
- Morton, C. M., Dayanandan, S., and Dissanayake, D., (1999) Phylogeny and biosystematics of *Pseudomonotes* (Dipeterocarpaceae) based on molecular and morphological data. Pl. Syst. Evol. **216**, 197–205.
- Neuhaus, H., and Link, G., (1987) The chloroplast tRNA^{Lys}(UUU) gene from mustard (Sinapis alba) contains a class II intron potentially coding for a maturase-related polypeptide. Curr. Gene. **11**, 251–257.
- Oginuma, K., Kono, Y., and Tobe, H., (1999) Embryology of Vateriopsis (Dipterocarpaceae) and relationships of the family. In: Abstract for the XVI International Botanical Congress, http://www.biologie.uni-hamburg.de/bonline/ibc99/ibc/abstracts/ listen/abstr acts/3183.html.
- Ooi, K., Endo, Y., Yokoyama, J., and Murakami, N., (1995) Useful primer designs to amplify DNA fragments of the plastid gene *matK* from Angiosperm plants. J. Jap. Bot. **70**, 328– 331.
- Parameswaran, N., and Gotwald, H., (1979) Problematic taxa in the Dipterocarpaceae. Their Anatomy and Taxonomy. In: Diptérocarpacées: Taxonomie-Phylogénie-Ecologie (ed.: G. Maury-Lechon), pp. 69–75. Mémoires du Muséum National d'Histoire Naturelle: First International Round Table on Dipterocarpaceae, Editions du Muséum, Paris.
- Saitou, N., and Nei, M., (1987) The neighbor-joining method: A new method for reconstructing phylogenetic trees. Mol. Biol. Evol. 4, 406–425.
- Symington, C. F., (1943) Foresters, manual of Dipterocarps. Malayan forest records No. 16, Penerbit Universiti Malaya, Kuala Lumpur, Malaysia, pp. 244.
- Taberlet, P., Gielly, L., Pautou, G., and Bouvet, J., (1991) Universal primers for amplification of three non-coding regions of chloroplast DNA. Pl. Mol. Biol. **17**, 1105–1109.
- Thompson, J. D., Gibson, T. J., Plewniak, F., and Higgins, D. G.,

(1997) The ClustalX windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. Nucleic Acids Res. **24**, 4876–4882.

Tsumura, Y., Kawahara, T., Wickneswari, R., and Yoshimura,

K., (1996) Molecular phylogeny of Dipterocarpaceae in Southeast Asia using RFLP of PCR-amplified chloroplast genes. Theor. Appl. Genet. **93**, 22–29.